

Della Volpe et al

```
library(Seurat)
library(clusterProfiler)
library(dplyr)
library(viridis)
library(ggplot2)
library(grid)
library(gridExtra)
library(reshape2)
library(scales)
library(RColorBrewer)
library(ComplexHeatmap)
library(circlize)
library(stringr)
library(ggalluvial)
library(ggrepel)
library(openxlsx)
library(tidyr)
library(ggalluvial)
library(gtools)
library(cowplot)
#devtools::install_github("psyteachr/introdataviz")
library('introdataviz')
library(ggpubr)

full_obj <- readRDS(paste(wdir,"Full_final.rds", sep = "/")) #resolution 2
```

Figure 4 b - Umap annotated clusters

```
mycols <- c('HSC_MPP' = "lightblue1", 'MPP' = 'skyblue2',
'LMPP'="skyblue3", 'LyP'="sandybrown",
'GMP_1' = "lightgreen", 'GMP_2'="#0CB702", 'GMP_granulocyte' =
"olivedrab4",
'MDP' = "#8494FF", 'ProB' = "#E68613", 'MEP'="lightpink",
'MEMBP_1' = "indianred1", 'MEMBP_2'="indianred3", 'EryP'="indianred4",
'EBMP' = "#FF68A1", "MEP_Baso_Mast"="#CD9600" )

Idents(full_obj) <- full_obj$Celltype
p <- DimPlot(full_obj, pt.size = 0.45, cols=mycols)
px <- LabelClusters(p, id = "ident", repel=T, color = "black",size=5)
#+theme_void()
theme(axis.text=element_text(size=10),
axis.line = element_line(colour = "black", size=0.7),
axis.title=element_text(size=22),
```

```
axis.text.x = element_text(hjust=1,size=10),
axis.text.y = element_text(size = 10),
panel.grid = element_blank(),
panel.border = element_blank(),
text = element_text( size= 12)) +
xlab('UMAP 1') + ylab('UMAP 2')
```

```
#svg(paste(out_dir,"Paper_plots", "umap_.svg", sep='/'), width =13, height = 10,
pointsize=12)
```

```
#px
```

```
#dev.off()
```

```
px
```

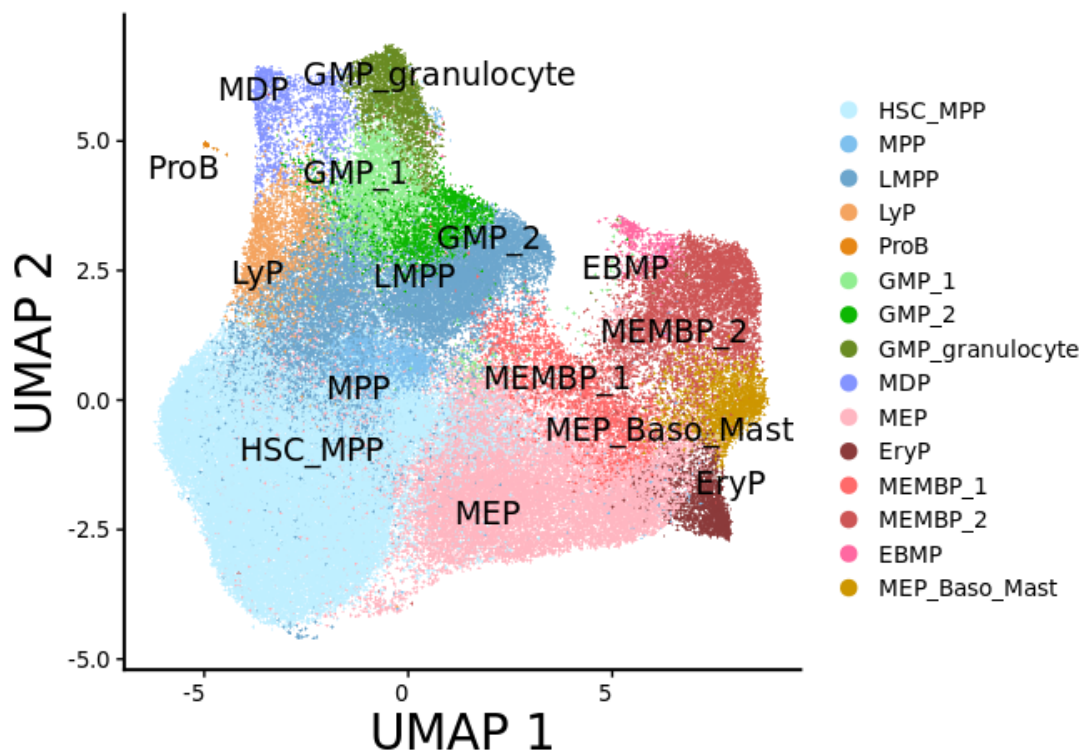


Figure 4 c - Barplot full object cell type cluster composition

```
table_cell_comp <- full_obj@meta.data %>%
  group_by(Celltype, RNA_pop) %>%
  count() %>%
  group_by(RNA_pop) %>%
  mutate(percent=100*n/sum(n)) %>%
  ungroup()
```

```

table_cell_comp$treatment <- ifelse(table_cell_comp$RNA_pop == 'UT_DMSO_pop', 'Not
edited',
                                ifelse(table_cell_comp$RNA_pop == 'UT_p38i_pop', 'Not
edited',
                                ifelse(table_cell_comp$RNA_pop == 'HS_DMSO_pop', 'HS
Cas9',
                                ifelse(table_cell_comp$RNA_pop == 'HS_p38i_pop', 'HS
Cas9',
                                ifelse(table_cell_comp$RNA_pop == 'GE_DMSO_pop_GFP-',
'GFP-',
ifelse(table_cell_comp$RNA_pop == 'GE_p38i_pop_GFP-', 'GFP-',
ifelse(table_cell_comp$RNA_pop == 'GE_DMSO_pop_GFP+',
'GFP+',
ifelse(table_cell_comp$RNA_pop == 'GE_p38i_pop_GFP+', 'GFP+',
'ND' )))))))

table_cell_comp$treatment <- factor(table_cell_comp$treatment, levels=c(
  "Not edited", "HS Cas9", "GFP-", "GFP+"
))

table_cell_comp$Sample_name <- ifelse(table_cell_comp$RNA_pop ==
'UT_DMSO_pop', 'DMSO',
                                ifelse(table_cell_comp$RNA_pop ==
'UT_p38i_pop', 'p38i',
                                ifelse(table_cell_comp$RNA_pop ==
'HS_DMSO_pop', 'DMSO',
                                ifelse(table_cell_comp$RNA_pop == 'HS_p38i_pop',
'p38i',
                                ifelse(table_cell_comp$RNA_pop == 'GE_DMSO_pop_GFP-
', 'DMSO',
                                ifelse(table_cell_comp$RNA_pop == 'GE_p38i_pop_GFP-
', 'p38i',
ifelse(table_cell_comp$RNA_pop == 'GE_DMSO_pop_GFP+', 'DMSO',
ifelse(table_cell_comp$RNA_pop ==
'GE_p38i_pop_GFP+', 'p38i',
'ND' )))))))

## save table
d_list <- list()
d_num <- dcast(table_cell_comp[,c(1,2,3)], RNA_pop ~ Celltype)
d_perc <- dcast(table_cell_comp[,c(1,2,4)], RNA_pop ~ Celltype)
d_list[["Number_Cells_CC_cluster"]] <- d_num
d_list[["Perc_Cells_CC_cluster"]] <- d_perc
## save perc and number
#out_dir <- 'Full/Paper_plots/tables'
#write.xlsx(x = d_list, file = paste(out_dir, 'Celltype_full_cell_perc.xlsx', sep =
"/"), rowNames = FALSE)

table_cell_comp$Celltype <- factor(table_cell_comp$Celltype, levels =

```

```

rev(levels(table_cell_comp$Celltype)))

p1 <- table_cell_comp %>%
  ggplot(aes(x=Sample_name,y=percent, fill=Celltype)) +
  geom_col(aes(fill = Celltype), width = .5, color = "black")+
  #ggtitle("Percentage of cell cycle phases per cluster") +
  scale_fill_manual(name = '', values = mycols) +
  theme_classic() +
  scale_y_continuous(name = '% of cells (Alive + CD90+)', expand = c(0.01,0)) +
  geom_flow(aes(alluvium = Celltype),min.y= -1, alpha= .9,
            lty = 2, fill = "white", color = "black",
            curve_type = "linear",
            width = .5) +
  facet_wrap(~treatment,nrow=1, strip.position = "bottom") +
  theme(
    legend.position = 'right',
    plot.title = element_text(hjust = 0.5),
    text = element_text(size = 16),
    panel.grid.major = element_blank(),
    panel.grid.minor = element_blank(),
    axis.title.x = element_blank(),
    plot.margin = margin(t = 20, r = 10, b = 10, l = 10, unit = 'pt'),
    axis.line = element_line(colour = "black",
                             size = 0.3, linetype = "solid"),
    axis.text.x = element_text(size=12, color='black'), # angle=0, hjust=1,
    axis.text.y = element_text(size=12, color='black'),
    strip.placement = "outside")

#svg(paste(out_dir,"barplot_full_celltype_composition.svg", sep='/'), width = 8,
height = 4, pointsize=12)
#p1
#dev.off()
p1

```

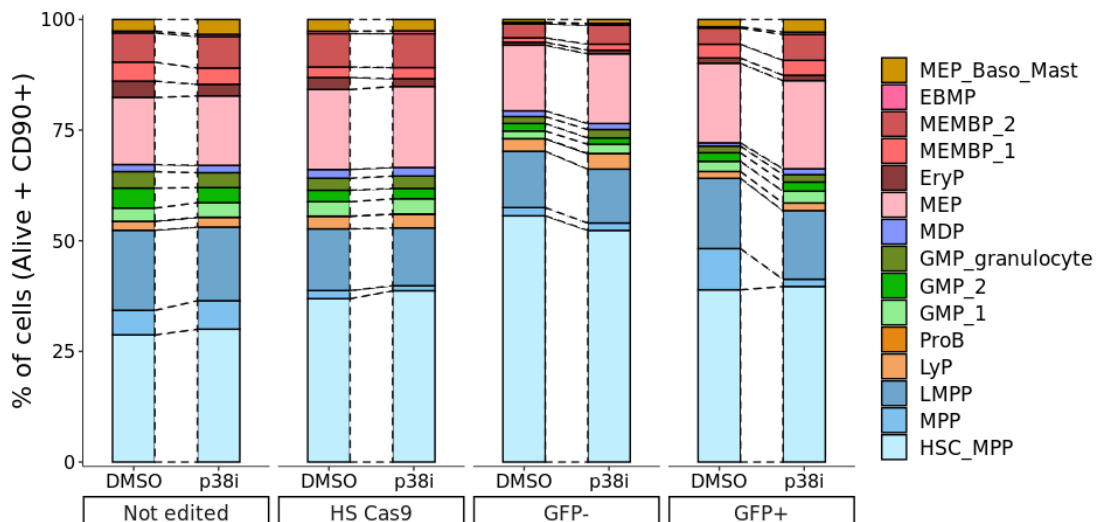


Figure 4 d - Barplot cell type cluster composition in sorted population CD34+CD133+CD90+CD45RA-

```
table_cell_comp <- full_obj@meta.data %>%
  group_by(Celltype, RNA_hashtag) %>%
  count() %>%
  group_by(RNA_hashtag) %>%
  mutate(percent=100*n/sum(n)) %>%
  ungroup()

table_cell_comp$type <- unlist(lapply(str_split(table_cell_comp$RNA_hashtag,
  '_'), "[", 3))
table_cell_comp <- table_cell_comp[table_cell_comp$type == 'CD90+',]
table_cell_comp$treatment <- ifelse(table_cell_comp$RNA_hashtag ==
  "UT_p38i_CD90+", 'Not edited',
  ifelse(table_cell_comp$RNA_hashtag ==
  "UT_DMSO_CD90+", 'Not edited',
  ifelse(table_cell_comp$RNA_hashtag ==
  "HS_DMSO_CD90+", 'HS Cas9',
  ifelse(table_cell_comp$RNA_hashtag ==
  "HS_p38i_CD90+", 'HS Cas9',
  ifelse(table_cell_comp$RNA_hashtag ==
  "GE_DMSO_CD90+_GFP-", 'GFP-',
  ifelse(table_cell_comp$RNA_hashtag ==
  "GE_p38i_CD90+_GFP-", 'GFP-',
  ifelse(table_cell_comp$RNA_hashtag ==
  "GE_DMSO_CD90+_GFP+", 'GFP+',
  ifelse(table_cell_comp$RNA_hashtag ==
  "GE_p38i_CD90+_GFP+", 'GFP+',
  'ND' )))))))

table_cell_comp$treatment <- factor(table_cell_comp$treatment, levels=c(
  "Not edited", "HS Cas9", "GFP-", "GFP+"
))

table_cell_comp$Sample_name <- ifelse(table_cell_comp$RNA_hashtag ==
  "UT_p38i_CD90+", 'p38i',
  ifelse(table_cell_comp$RNA_hashtag ==
  "UT_DMSO_CD90+", 'DMSO',
  ifelse(table_cell_comp$RNA_hashtag ==
  "HS_DMSO_CD90+", 'DMSO',
  ifelse(table_cell_comp$RNA_hashtag ==
  "HS_p38i_CD90+", 'p38i',
  ifelse(table_cell_comp$RNA_hashtag ==
  "GE_DMSO_CD90+_GFP-", 'DMSO',
  ifelse(table_cell_comp$RNA_hashtag ==
  "GE_p38i_CD90+_GFP-", 'p38i',
```

```

        ifelse(table_cell_comp$RNA_hashtag ==
"GE_DMSO_CD90+_GFP+", 'DMSO',
        ifelse(table_cell_comp$RNA_hashtag ==
"GE_p38i_CD90+_GFP+", 'p38i',
        'ND' )))))))

## save table
d_list <- list()
d_num <- dcast(table_cell_comp[,c(1,2,3)], RNA_hashtag ~ Celltype)
d_perc <- dcast(table_cell_comp[,c(1,2,4)], RNA_hashtag ~ Celltype)
d_list[["Number_Cells_CC_cluster"]] <- d_num
d_list[["Perc_Cells_CC_cluster"]] <- d_perc
## save perc and number

#write.xlsx(x =d_list, file = paste(out_dir, 'Celltype_CD90_cell_perc.xlsx', sep =
"/"), rowNames = FALSE)

table_cell_comp_CD90 <- table_cell_comp
table_cell_comp_CD90$Celltype <- factor(table_cell_comp_CD90$Celltype, levels =
rev(levels(table_cell_comp_CD90$Celltype)))

p1 <- table_cell_comp_CD90 %>%
  ggplot(aes(x=Sample_name,y=percent, fill=Celltype)) +
  geom_col(aes(fill = Celltype), width = .5, color = "black")+
  #ggtitle("Percentage of cell cycle phases per cluster") +
  scale_fill_manual(name = '', values = mycols) +
  theme_classic() +
  scale_y_continuous(name = '% of CD90+ cells', expand = c(0.01,0)) +
  geom_flow(aes(alluvium = Celltype),min.y= -1, alpha= .9,
            lty = 2, fill = "white", color = "black",
            curve_type = "linear",
            width = .5) +
  facet_wrap(~treatment,nrow=1, strip.position = "bottom") +
  theme(
    legend.position = 'right',
    plot.title = element_text(hjust = 0.5),
    text = element_text(size = 16),
    panel.grid.major = element_blank(),
    panel.grid.minor = element_blank(),
    axis.title.x = element_blank(),
    plot.margin = margin(t = 20, r = 10, b = 10, l = 10, unit = 'pt'),
    axis.line = element_line(colour = "black",
                             size = 0.3, linetype = "solid"),
    axis.text.x = element_text(size=12, color='black'), # angle=0, hjust=1,
    axis.text.y = element_text(size=12, color='black'),
    strip.placement = "outside")

#svg(paste(out_dir,"barplot_cd90+_celltype_composition.svg", sep='/'), width =8,

```

```
height = 4, pointsize=12)
#p1
#dev.off()
p1
```

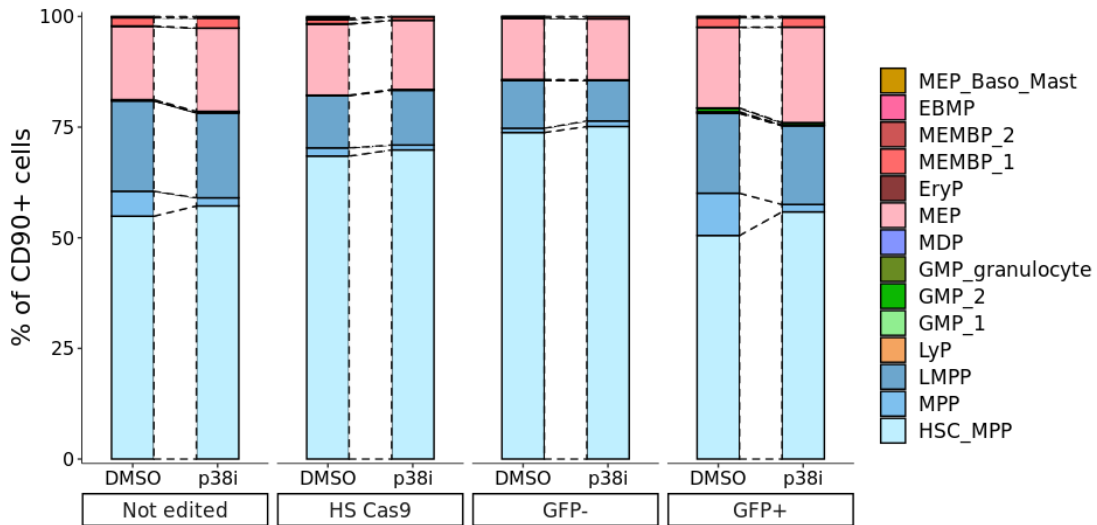


Figure 4 h - GSEA hsc signatures from literature Laurenti et al.

```
lset <- list('hsc' = c('RodriguezFraticelli_2020.gmt', 'Garcia-Prat2021.gmt'))

c <- list("UT_p38i_pop_vs_UT_DMSO_pop" = c("UT_p38i_pop", "UT_DMSO_pop"),
         "HS_p38i_pop_vs_HS_DMSO_pop" = c("HS_p38i_pop", "HS_DMSO_pop"),
         "GE_DMSO_pop_GFP+_vs_GE_DMSO_pop_GFP-" =
c("GE_DMSO_pop_GFP+", "GE_DMSO_pop_GFP-"), ## h3 vs h3
         "GE_p38i_pop_GFP+_vs_GE_DMSO_pop_GFP+" =
c("GE_p38i_pop_GFP+", "GE_DMSO_pop_GFP+"), ## h4 vs h3
         "GE_p38i_pop_GFP-_vs_GE_DMSO_pop_GFP-" = c("GE_p38i_pop_GFP-",
         "GE_DMSO_pop_GFP-"),
         "GE_DMSO_pop_GFP+_vs_UT_DMSO_pop" = c("GE_DMSO_pop_GFP+", "UT_DMSO_pop"),
## h3 vs h3
         "GE_DMSO_pop_GFP-_vs_UT_DMSO_pop" = c("GE_DMSO_pop_GFP-", "UT_DMSO_pop"),
         "GE_p38i_pop_GFP+_vs_UT_DMSO_pop" = c("GE_p38i_pop_GFP+", "UT_DMSO_pop"),
         "GE_p38i_pop_GFP-_vs_UT_DMSO_pop" = c("GE_p38i_pop_GFP-", "UT_DMSO_pop"))
## h3 vs h3

x <- c('HSC_MPP')

comparisons <- expand.grid(x, names(c))
comp <- paste0(comparisons$Var1, '_', comparisons$Var2)
list_comp <- c()
gsea_res <- list()
```

```

out_dir <- 'Full/DGE_analysis/GSEA_alivecd90_together_HSC'
marker_dir <- 'Full/DGE_analysis/DGE_alivecd90_together/tables'
for (comp in comp) { #for each comparisons within cluster
  for(nn in names(lset)){ #gmt list
    lset_name = lset[[nn]] ## get lists
    fi <- paste(out_dir, paste0(nn, '_', comp, "_GSEA.xlsx"), sep = "/")

    if (file.exists(fi)) { next }
    for (gset in lset_name) { #get specific gmt file

      t2gene <- read.gmt(gmtfile = paste0("reference/GSEA/", gset))

      ## check if file exist
      f <- paste(marker_dir, paste0(comp, '.txt'), sep = '/')
      if (file.exists(f)) {
        markers <- read.table(f)

        genelist <- markers$avg_log2FC
        names(genelist) <- rownames(markers) # $row.names
        # genelist <- na.omit(genelist)
        genelist <- sort(genelist, decreasing = T)

        print(paste0("Running GSEA ", comp, " - ", gset, '-', nn ))

        ## do not run again
        #gsea_res[[nn]][[gset]][[comp]] <- GSEA(genelist, TERM2GENE = t2gene,
        #                                     maxGSsize = 1000, minGSsize =
10, seed = 123,
        #                                     verbose = FALSE,
        #                                     pvalueCutoff = 1)
      }

      if (length(gsea_res[[nn]][[gset]][[comp]]) > 0){
        gsea_res[[nn]][[gset]][[comp]]@result['comparisons'] <- comp
        #write.xlsx(x = gsea_res[[nn]][[gset]][[comp]], file = paste(out_dir,
paste0(nn, '_', comp, "_GSEA.xlsx"), sep = "/"), rowNames = TRUE)
      }
    }
  }
}

#saveRDS(gsea_res, file = paste(out_dir, 'gsea_deg_gseaplot_HSC_mouse_human.rds',
sep = "/"))
gsea_res <- readRDS(paste(out_dir,
'gsea_deg_gseaplot_HSC_mouse_human.rds', sep = '/'))

only_hsc <- c('HSC_MPP')

```



```

c <- list("UT_p38i_pop_vs_UT_DMSO_pop" = c("UT_p38i_pop", "UT_DMSO_pop"),
         "HS_p38i_pop_vs_HS_DMSO_pop" = c("HS_p38i_pop", "HS_DMSO_pop"),
         "GE_p38i_pop_GFP+_vs_GE_DMSO_pop_GFP+" =
c("GE_p38i_pop_GFP+", "GE_DMSO_pop_GFP+"),
         "GE_p38i_pop_GFP-_vs_GE_DMSO_pop_GFP-" = c("GE_p38i_pop_GFP-",
", "GE_DMSO_pop_GFP-"))

comparisons <- expand.grid(only_hsc, names(c))

hallmark.full <- data.frame()
comp <- paste0(comparisons$Var1, '_', comparisons$Var2)
for (comp in comp) {
  for(nn in names(lset)){ # m, arco-group for instance hema, senescence in gmt list
    lset_name = lset[[nn]]
    for (gset in lset_name) {
      df <- gsea_res[[nn]][[gset]][[comp]]@result
      df$Category <- gset
      df$Dataset <- comp
      if (nrow(hallmark.full) == 0) {
        hallmark.full <- df
      } else {
        hallmark.full <- rbind(hallmark.full, df)
      }
    }
  }
}
#####
#write.table(hallmark.full.filt, paste(out_dir, 'gsea_hsc_HUMAN_MOUSE.txt'))
hallmark.full.filt <- hallmark.full

hallmark.full.filt$p.adjust <- round(hallmark.full.filt$p.adjust)
hallmark.subset <- subset(hallmark.full.filt , p.adjust <= 0.1)

min0 <- min(hallmark.subset[!is.na(hallmark.subset$NES), 'NES'])
max0 <- max(hallmark.subset[!is.na(hallmark.subset$NES), 'NES'])

hallmark.subset$stars=stars.pval(hallmark.subset$pvalue)

hallmark.order <- hallmark.subset%>% group_by(ID) %>% summarise(Pos = sum(NES))
hallmark.order.terms <- hallmark.order[order(hallmark.order$Pos, decreasing =
TRUE), "ID", drop = FALSE]
hallmark.subset$ID <- factor(hallmark.subset$ID, levels = hallmark.order.terms$ID)

hallmark.subset$Dataset <- factor(hallmark.subset$Dataset,
levels=c("HSC_MPP_UT_p38i_pop_vs_UT_DMSO_pop",
"HSC_MPP_HS_p38i_pop_vs_HS_DMSO_pop",

```

```

"HSC_MPP_GE_p38i_pop_GFP-_vs_GE_DMSO_pop_GFP-",
"HSC_MPP_GE_p38i_pop_GFP+_vs_GE_DMSO_pop_GFP+") )

hallmark.subset$Dataset <- ifelse(hallmark.subset$Dataset ==
"HSC_MPP_UT_p38i_pop_vs_UT_DMSO_pop", 'Not edited',
      ifelse(hallmark.subset$Dataset
=="HSC_MPP_HS_p38i_pop_vs_HS_DMSO_pop", 'HS Cas9',
      ifelse(hallmark.subset$Dataset
=="HSC_MPP_GE_p38i_pop_GFP-_vs_GE_DMSO_pop_GFP-", "GFP-",
ifelse(hallmark.subset$Dataset == "HSC_MPP_GE_p38i_pop_GFP+_vs_GE_DMSO_pop_GFP+",
"GFP+", 'none' ))))

hallmark.subset$Dataset <- factor(hallmark.subset$Dataset, levels=rev(c('Not
edited',
                                                                    'HS Cas9',
                                                                    "GFP-",
                                                                    "GFP+") ))

hallmark.subset$ID <- gsub('_', ' ', hallmark.subset$ID)
hallmark.subset$ID <- str_wrap(hallmark.subset$ID ,width = 30)
name_list <- list('Lauridsen et al 2019'=c("RACFP DIM Lauridsen et al"),
      'Rodriguez et al 2020'=c("HSC1 cluster","HSC2 cluster","HSC3
cluster","HSC4 cluster",
      "Low-output Rodriguez et al","High-output Rodriguez et al", "Mk-
bias Rodriguez et al","Serial-engraftment Rodriguez\net al", "Multilineage
Rodriguez et al"),
      'Pietras et al 2014' = c("HSC Pietras et al"),
      'Stemscore Giladi et al' = c("Stemscore Giladi et al"),
      'Wilson et al' = c("SuMO Wilson et al", "MolO Wilson et al"),
      'Cabezas-Wallscheid et al' = c("dHSC aHSC single-cell\nCabezas-
Wallscheid et al","HSC vs MPP4 Cabezas-Wallscheid\net al","dHSC vs aHSC\nCabezas-
Wallscheid et al"))

hallmark.subset$Name <- ifelse(hallmark.subset$ID %in% name_list$`Lauridsen et al
2019`, 'Lauridsen et al 2019',
      ifelse(hallmark.subset$ID %in% name_list$`Rodriguez et al
2020`, 'Rodriguez et al 2020',
      ifelse(hallmark.subset$ID %in% name_list$`Pietras et al
2014`, 'Pietras et al 2014',
      ifelse(hallmark.subset$ID %in% name_list$`Stemscore Giladi
et al`, "Stemscore Giladi et al",
      ifelse(hallmark.subset$ID %in% name_list$`Wilson et al`,
"Wilson et al",
      ifelse(hallmark.subset$ID %in% name_list$`Cabezas-
Wallscheid et al`, 'Cabezas-Wallscheid et al', 'Garcia-Prat' ))))))

# exclude singatures
hallmark.subset <- hallmark.subset[!hallmark.subset$ID %in% c("HSC Pietras et al",
"Mk-bias Rodriguez et al","Serial-engraftment Rodriguez\net al", "Multilineage
Rodriguez et al"),]

```


Figure 4 e - GSEA hallmark & GO BP

```
hallmark.subset <- read.table('Full/Paper_plots/tables/Main_Figure_4e.txt' )

min0 <- min(hallmark.subset[!is.na(hallmark.subset$NES), 'NES'])
max0 <- max(hallmark.subset[!is.na(hallmark.subset$NES), 'NES'])

hallmark.subset$stars=stars.pval(hallmark.subset$pvalue)

## adjust name too Long
val <- "regulation of transcription involved in G1/S transition of mitotic cell
cycle (GO:0000083)"
valx <- "reg. transcr. involved in G1/S transition (GO:0000083)"
hallmark.subset$ID <- ifelse(hallmark.subset$ID == val, valx, hallmark.subset$ID)
hallmark.subset$Name <- ifelse(hallmark.subset$Category == 'go',
sapply(strsplit(as.character(hallmark.subset$ID), "\\("), `[`, 1),
as.character(hallmark.subset$ID))
hallmark.subset$ID <- str_wrap(hallmark.subset$ID ,width = 45)
hallmark.subset$celltype <- gsub('_', ' ', hallmark.subset$celltype)
hallmark.subset$celltype <- ifelse(hallmark.subset$celltype == 'MEP Baso Mast',
'MEP Baso-Mast', hallmark.subset$celltype)
#hallmark.subset$celltype <- ifelse(nchar(hallmark.subset$celltype) > 7,
str_wrap(hallmark.subset$celltype ,width = 6), hallmark.subset$celltype)

hallmark.subset$celltype <- factor(hallmark.subset$celltype, levels =
c("HSC MPP", 'MPP', "LMPP", "GMP 1",
"GMP 2", "GMP\ngranulocyte",
"MDP", "MEP", "EryP", "MEMBP 1", "MEMBP 2",
"MEP\nBaso-Mast", "LyP", "ProB", "EBMP" ))

hallmark.order <- hallmark.subset%>% group_by(ID) %>% summarise(Pos = sum(NES))
hallmark.order.terms <- hallmark.order[order(hallmark.order$Pos, decreasing =
TRUE), "ID", drop = FALSE]
hallmark.subset$ID <- factor(hallmark.subset$ID, levels = hallmark.order.terms$ID)

hallmark.subset$Dataset <- factor(hallmark.subset$Dataset,
levels=c("UT_p38i_pop_vs_UT_DMSO_pop",
"HS_p38i_pop_vs_HS_DMSO_pop",
"GE_p38i_pop_GFPneg_vs_GE_DMSO_pop_GFPneg",
"GE_p38i_pop_GFPpos_vs_GE_DMSO_pop_GFPpos"))

hallmark.subset$Dataset <- ifelse(hallmark.subset$Dataset ==
"UT_p38i_pop_vs_UT_DMSO_pop", 'Not edited',
ifelse(hallmark.subset$Dataset
```

```

=="HS_p38i_pop_vs_HS_DMSO_pop", 'HS Cas9',
      ifelse(hallmark.subset$Dataset
=="GE_p38i_pop_GFPneg_vs_GE_DMSO_pop_GFPneg", "GFP-",
      ifelse(hallmark.subset$Dataset
=="GE_p38i_pop_GFPpos_vs_GE_DMSO_pop_GFPpos", "GFP+", 'none' ))))

hallmark.subset$Dataset <- factor(hallmark.subset$Dataset, levels=c('Not edited',
                                                                    'HS Cas9',
                                                                    "GFP-",
                                                                    "GFP+" )

hallmark.subset <- hallmark.subset[hallmark.subset$Category != 'p38_signature',]
p <- ggplot(data = hallmark.subset, mapping = aes(x = Dataset, y = reorder(Name, -
NES))) +
  geom_tile(aes(fill=NES, group = celltype), width=0.80, height=0.80)+
  facet_grid(Category ~ celltype , scales = "free_y", space = "free", )+
  theme(text= element_text(size=22,color='black'),
        axis.text.x = element_text(angle = 90, vjust = 0.5, hjust=1,
color='black', size =12),
        axis.text.y = element_text(color='black', size =16),
        plot.title = element_text(hjust = 0), plot.subtitle = element_text(hjust =
0),
        axis.ticks = element_blank(), axis.text = element_text(size = 8),
axis.title = element_blank(),
        panel.grid.major = element_blank(), panel.grid.minor = element_blank(),
        panel.background = element_rect(fill = "gray95", color='black'),
        strip.background = element_rect(colour="white",
        fill="white",size=0.5),
        strip.text = element_text(color='black',face = 'bold', size = 11),
        legend.position = 'top', legend.direction = 'horizontal') +
  scale_fill_gradientn(colours =
colorRampPalette(rev(brewer.pal(11,"RdBu")))(100),
        limits = c(min0,max0),
        na.value = "grey")+ ggtitle('p38i vs DMSO')

#wdir='Full/DGE_analysis'
#svg(paste(wdir,"HM_categories_v4.svg",sep = '/'),width=22,
# height=11, pointsize=12)
#p
#dev.off()
p

```

p38i vs DMSO

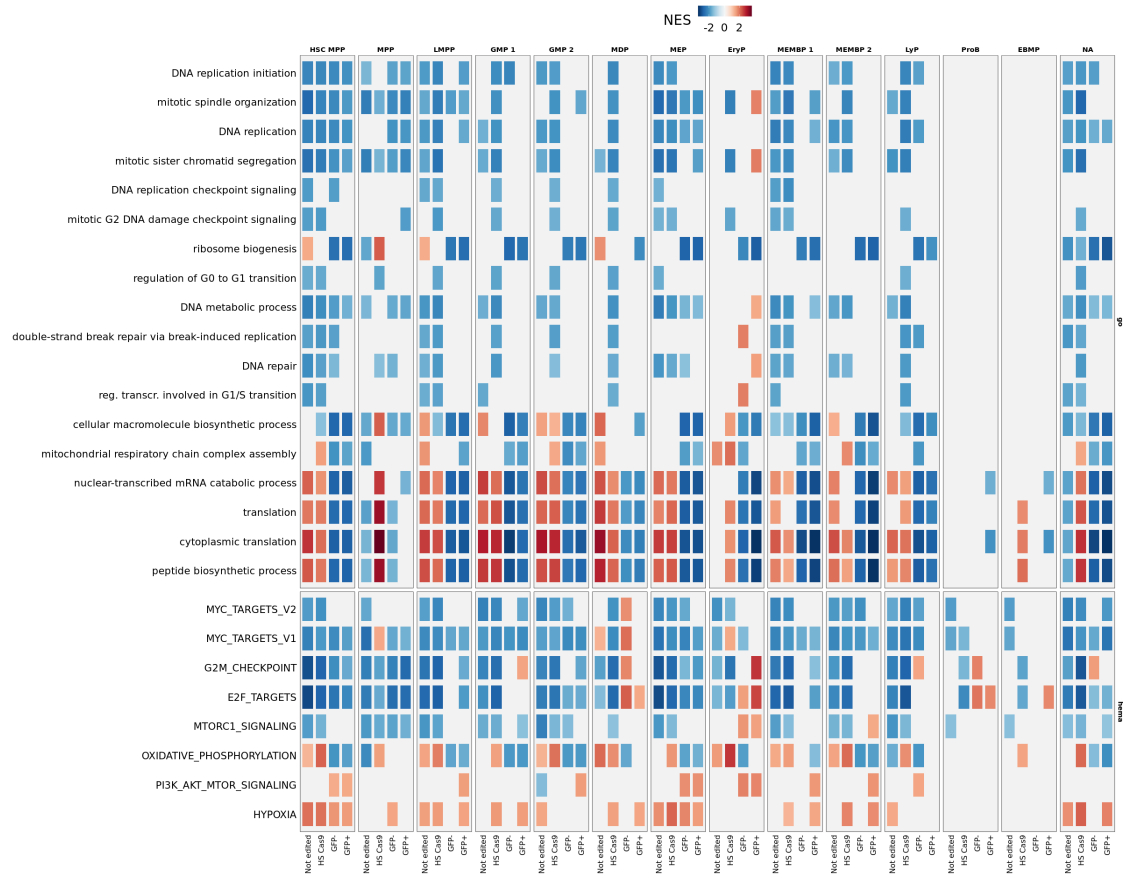


Figure 4 f - cell cycle full object & per cluster

```
full_obj$RNA_pop <- factor( full_obj$RNA_pop,
levels=c("UT_DMSO_pop", "UT_p38i_pop",
"HS_DMSO_pop",
"HS_p38i_pop",
"GE_DMSO_pop_GFP+", "GE_p38i_pop_GFP+",
"GE_DMSO_pop_GFP-",
"GE_p38i_pop_GFP-"))
table_cell_cycle <- full_obj@meta.data %>%
  group_by(RNA_pop, Phase) %>%
  count() %>%
  group_by(RNA_pop) %>%
  mutate(percent=100*n/sum(n)) %>%
  ungroup()

d_list <- list()
d_num <- dcast(table_cell_cycle[,c(1,2,3)], Phase ~ RNA_pop)
d_perc <- dcast(table_cell_cycle[,c(1,2,4)], Phase ~ RNA_pop)
d_list[["Number_Cells_CC_treatment"]] <- d_num
```

```

d_list[["Perc_Cells_CC_treatment"]] <- d_perc

#write.xlsx(x = d_list, file = paste(out_dir, 'Cellcycle_treatment.xlsx', sep =
"/"), rowNames = FALSE)

custom_colors <- list()
custom_colors$cell_cycle <- setNames(
  c("blue", "red", "gray60"), #'#7f8c8d',
  c('G1', 'S', 'G2M') # '-'
)

table_cell_cycle$Phase <- factor(table_cell_cycle$Phase, levels =
c("G2M", "S", "G1"))
table_cell_cycle$treatment <- ifelse(table_cell_cycle$RNA_pop ==
'UT_DMSO_pop', 'Not edited',
ifelse(table_cell_cycle$RNA_pop ==
'UT_p38i_pop', 'Not edited',
ifelse(table_cell_cycle$RNA_pop == 'HS_DMSO_pop', 'HS
Cas9',
ifelse(table_cell_cycle$RNA_pop == 'HS_p38i_pop',
'HS Cas9',
', 'GFP-',
ifelse(table_cell_cycle$RNA_pop == 'GE_DMSO_pop_GFP-',
', 'GFP-',
ifelse(table_cell_cycle$RNA_pop == 'GE_p38i_pop_GFP-',
'GE_DMSO_pop_GFP+', 'GFP+',
ifelse(table_cell_cycle$RNA_pop ==
'GE_p38i_pop_GFP+', 'GFP+', 'ND' )))))))

table_cell_cycle$treatment <- factor(table_cell_cycle$treatment, levels=c(
  "Not edited", "HS Cas9", "GFP-", "GFP+"))

table_cell_cycle$Sample_name <- ifelse(table_cell_cycle$RNA_pop ==
'UT_DMSO_pop', 'DMSO',
ifelse(table_cell_cycle$RNA_pop ==
'UT_p38i_pop', 'p38i',
ifelse(table_cell_cycle$RNA_pop ==
'HS_DMSO_pop', 'DMSO',
ifelse(table_cell_cycle$RNA_pop == 'HS_p38i_pop',
'p38i',
ifelse(table_cell_cycle$RNA_pop ==
'GE_DMSO_pop_GFP-', 'DMSO',
ifelse(table_cell_cycle$RNA_pop ==
'GE_p38i_pop_GFP-', 'p38i',
ifelse(table_cell_cycle$RNA_pop ==
'GE_DMSO_pop_GFP+', 'DMSO',
ifelse(table_cell_cycle$RNA_pop ==
'GE_p38i_pop_GFP+', 'p38i',
'ND' )))))))

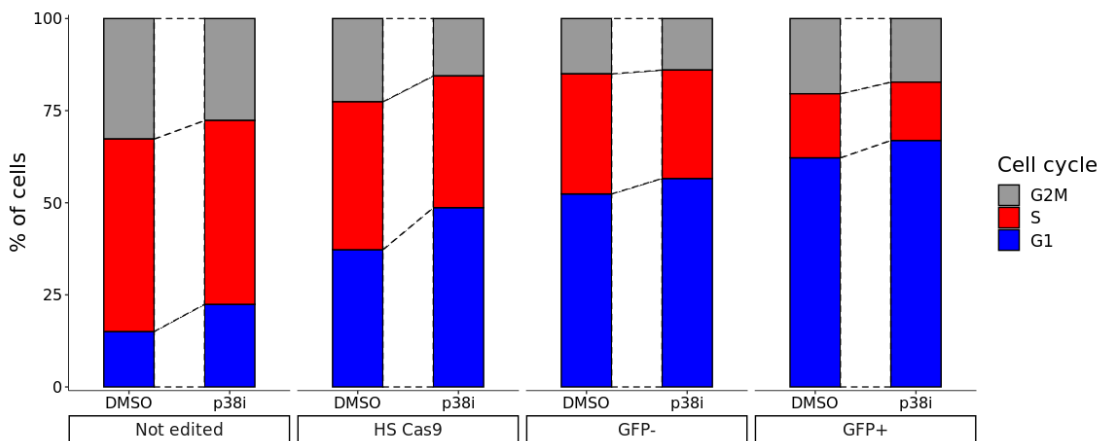
```

```

p1 <- table_cell_cycle %>%
  ggplot(aes(x=Sample_name,y=percent, fill=Phase)) +
  geom_col(aes(fill = Phase), width = .5, color = "black")+
  #ggtitle("Percentage of cell cycle phases per cluster") +
  scale_fill_manual(name = 'Cell cycle', values = custom_colors$cell_cycle) +
  theme_classic() +
  scale_y_continuous(name = '% of cells', expand = c(0.01,0)) +
  geom_flow(aes(alluvium = Phase),min.y= -1, alpha= .9,
            lty = 2, fill = "white", color = "black",
            curve_type = "linear",
            width = .5) +
  facet_wrap(~treatment,nrow=1, strip.position = "bottom") +
  theme(
    legend.position = 'right',
    plot.title = element_text(hjust = 0.5),
    text = element_text(size = 16),
    panel.grid.major = element_blank(),
    panel.grid.minor = element_blank(),
    axis.title.x = element_blank(),
    plot.margin = margin(t = 20, r = 10, b = 10, l = 10, unit = 'pt'),
    axis.line = element_line(colour = "black",
                             size = 0.3, linetype = "solid"),
    axis.text.x = element_text(size=12, color='black'), # angle=0, hjust=1,
    axis.text.y = element_text(size=12, color='black'),
    strip.placement = "outside",
  )

#svg(paste(out_dir,"CC_treatment_perc_colorcode.svg", sep='/'), width =8, height =
4, pointsize=12)
#
#p1
# dev.off()
p1

```



```

#####
# cell cycle HSC cluster and all the others clusters
#####

```



```

table_cell_cycle <- full_obj@meta.data %>%
  group_by(Celltype, Phase, RNA_pop) %>%
  count() %>%
  group_by(RNA_pop, Celltype) %>%
  mutate(percent=100*n/sum(n)) %>%
  ungroup()

d_list <- list()
d_num <- dcast(table_cell_cycle[,c(1,2,3,4)], Celltype + Phase ~ RNA_pop )
d_perc <- dcast(table_cell_cycle[,c(1,2,3,5)], Celltype + Phase ~ RNA_pop)
d_list[["Number_Cells_CC_treatment"]] <- d_num
d_list[["Perc_Cells_CC_treatment"]] <- d_perc
out_dir <- 'Full/CellCycle'
#write.xlsx(x =d_List, file = paste(out_dir, 'Cellcycle_treatment_xcelltype.xlsx',
sep = "/"), rowNames = FALSE)

table_cell_cycle$Phase <- factor(table_cell_cycle$Phase, levels =
c("G2M", "S", "G1"))
table_cell_cycle$treatment <- ifelse(table_cell_cycle$RNA_pop ==
'UT_DMSO_pop', 'Not edited',
                                     ifelse(table_cell_cycle$RNA_pop ==
'UT_p38i_pop', 'Not edited',
                                             ifelse(table_cell_cycle$RNA_pop ==
'HS_DMSO_pop', 'HS Cas9',
                                                  ifelse(table_cell_cycle$RNA_pop
== 'HS_p38i_pop', 'HS Cas9',
                                                       ifelse(table_cell_cycle$RNA_pop ==
'GE_DMSO_pop_GFP-', 'GFP-',
                                                         ifelse(table_cell_cycle$RNA_pop ==
'GE_p38i_pop_GFP-', 'GFP-',
                                                             ifelse(table_cell_cycle$RNA_pop ==
'GE_DMSO_pop_GFP+', 'GFP+',
                                                                 ifelse(table_cell_cycle$RNA_pop
== 'GE_p38i_pop_GFP+', 'GFP+',
                                                                     'ND' )))))))

table_cell_cycle$treatment <- factor(table_cell_cycle$treatment, levels=c(
  "Not edited", "HS Cas9", "GFP-", "GFP+"
))

table_cell_cycle$Sample_name <- ifelse(table_cell_cycle$RNA_pop ==
'UT_DMSO_pop', 'DMSO',
                                       ifelse(table_cell_cycle$RNA_pop ==
'UT_p38i_pop', 'p38i',
                                               ifelse(table_cell_cycle$RNA_pop ==
'HS_DMSO_pop', 'DMSO',
                                                    ifelse(table_cell_cycle$RNA_pop == 'HS_p38i_pop',
'p38i',
                                                         ifelse(table_cell_cycle$RNA_pop ==
'GE_DMSO_pop_GFP-', 'DMSO',

```

```

        ifelse(table_cell_cycle$RNA_pop ==
'GE_p38i_pop_GFP-', 'p38i',
        ifelse(table_cell_cycle$RNA_pop ==
'GE_DMSO_pop_GFP+', 'DMSO',
        ifelse(table_cell_cycle$RNA_pop ==
'GE_p38i_pop_GFP+', 'p38i',
                                                    'ND'
))))))
p <- list()
for (n in unique(table_cell_cycle$Celltype)) {
  df <- data.frame(table_cell_cycle)
  dfx <- df[df$Celltype == as.character(n), ]
  dfx$roundfreq <- round(dfx$percent,2)

p[[n]] <- ggplot( dfx, aes(Sample_name, percent))+
  geom_col(aes(fill = Phase), width = .5, color = "black")+
  theme_classic() +
  #ggtitle("Percentage of cell cycle phases per cluster") +
  scale_fill_manual(name = '', values = custom_colors$cell_cycle) +
  theme_classic() +
  geom_flow(aes(alluvium = Phase),min.y= -1, alpha= .9,
            lty = 2, fill = "white", color = "black",
            curve_type = "linear",
            width = .5) + ggtitle(n)+
  scale_y_continuous(expand = c(0, 0))+
  facet_wrap(~treatment,nrow=1, strip.position = "bottom") +
  theme(
    legend.position = 'right',
    plot.title = element_text(hjust = 0.5),
    text = element_text(size = 16),
    panel.grid.major = element_blank(),
    panel.grid.minor = element_blank(),
    axis.title.x = element_blank(),
    plot.margin = margin(t = 20, r = 10, b = 10, l = 10, unit = 'pt'),
    axis.line = element_line(colour = "black",
                             size = 0.3, linetype = "solid"),
    axis.text.x = element_text(size=12, color='black'), # angle=0, hjust=1,
    axis.text.y = element_text(size=12, color='black'),
    #axis.title.y = element_blank(),
    strip.placement = "outside") + scale_y_continuous(name = '% of cells', expand
= c(0.01,0))
}

#barplot_pz <- grid.arrange(grobs =
List(p$HSC_MPP,p$MPP,p$LMPP,p$LyP,p$ProB,p$GMP_1,p$GMP_2,p$GMP_granulocyte,p$MDP,p
$MEP,p$EryP,p$MEMBP_1,p$MEMBP_2,p$EBMP,p$MEP_Baso_Mast),
#
      left = textGrob("", rot = 90, vjust = 1),ncol=4)

#Leg <- get_Legend(barplot_pz) ## extract the Legend before excluding it from plot
#p$Leg <- Leg

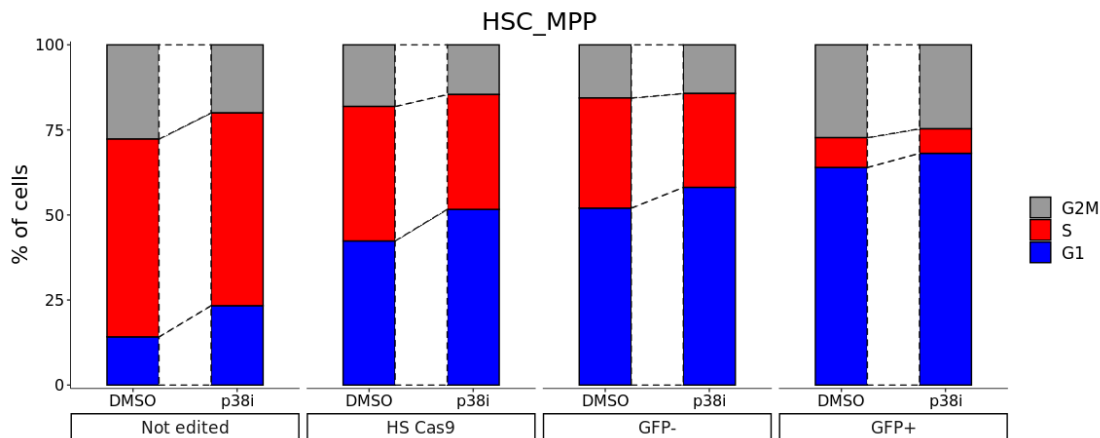
```

```

#svg(paste(out_dir,"CC_HSC_mpp_barplot_cellcycle_colorcoded.svg",sep =
'/' ),width=8,
# height=4, pointsize=12)

#dev.off()
grid.arrange(grobs = list(p$HSC_MPP),
             ncol=1)

```



Supplementary Figures

Supplementary Figure 4 d

```

#svg(paste(out_dir,"barplot_cellcycle_colorcoded.svg",sep = '/' ),width=10,
# height=60, pointsize=12)
#grid.arrange(grobs =
List(p$MPP,p$LMPP,p$LyP,p$ProB,p$GMP_1,p$GMP_2,p$GMP_granulocyte,p$MDP,p$MEP,p$Ery
P,p$MEMBP_1,p$MEMBP_2,p$EBMP,p$MEP_Baso_Mast),
# ncol=4)
#dev.off()

grid.arrange(grobs =
list(p$MPP,p$LMPP,p$LyP,p$ProB,p$GMP_1,p$GMP_2,p$GMP_granulocyte,p$MDP,p$MEP,p$Ery
P,p$MEMBP_1,p$MEMBP_2,p$EBMP,p$MEP_Baso_Mast), ncol=4)

```

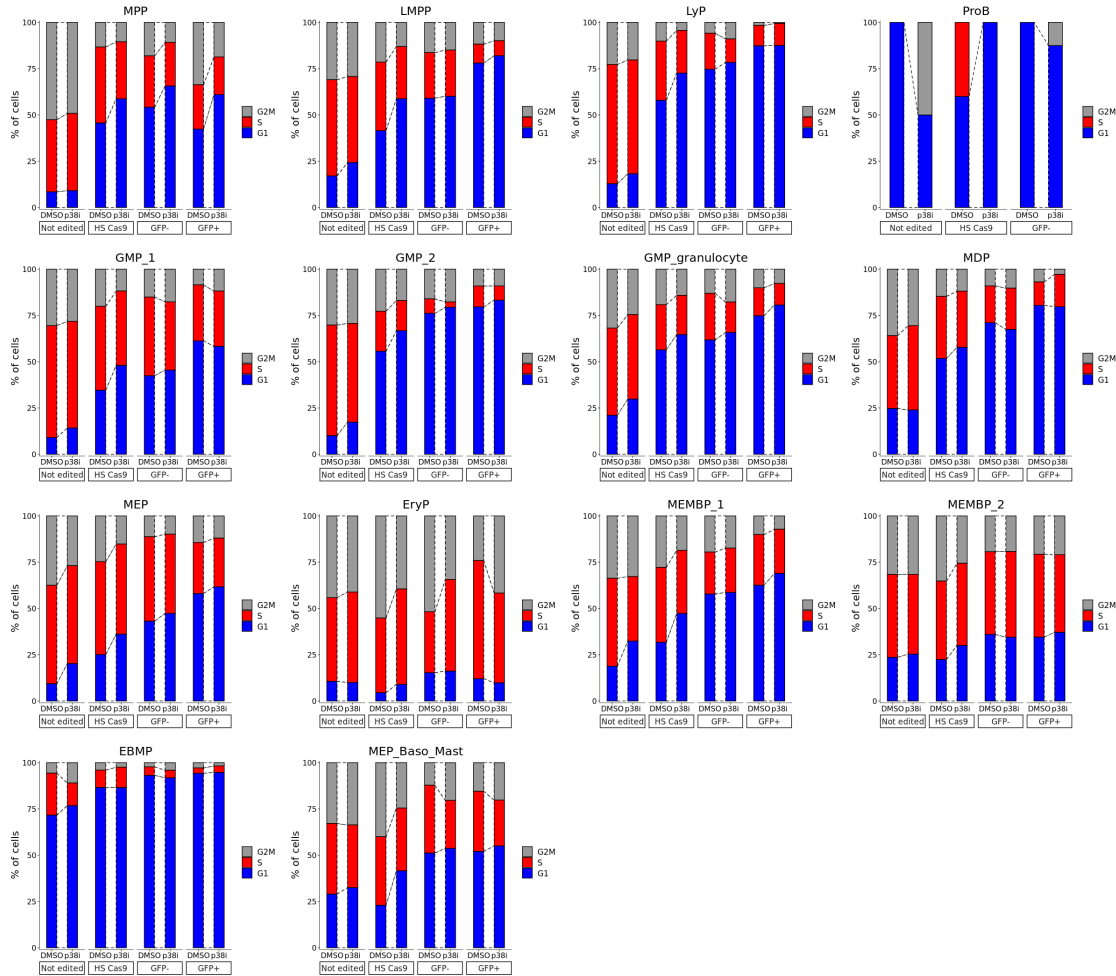


Figure 4 g - Lopes lists HSC-MPP

```
gmt.obj <- list(
  'Response_to_DNA_damage_replication_stress' =
  c('Plk1', 'Trex1', 'Atrip', 'Trp53', 'Tipin', 'Timeless', 'Rpa2', 'Rfdw3', 'Cdk1', 'Taok3',
    'Fzr1', 'Brcc3', 'Rad17', 'Babam2', 'Rad9a', 'Rad1', 'Cdc51', 'Etaa1',
    'Dtl', 'Chek1', 'Clspn', 'Hus1', 'Ticrr', 'Prpf19', 'Babam1', 'Tipr1', 'Abraxas1', 'Thoc1',
    'Topbp1', 'Donson', 'Chek2', 'Map3k20', 'Nek11', 'Mdm2', 'Pidd1',
    'Atr', 'Rhno1', 'Uimc1', 'Rbm38', 'Usp28', 'Atm', 'Trim39', 'Rad9b', 'Hus1b', 'Brsk1'),
  'DNA_replication_initiation' =
  c('Ccne1', 'Gmnc', 'Cdc45', 'Mcm10', 'Cdc6', 'Mcm2', 'Mcm6', 'Mcm7', 'Mcm4', 'Mcm5', 'Mcm3',
    'Cdt1', 'Noc31', 'Orc5', 'Orc3', 'Kat7', 'Orc1', 'Mcidas', 'Wrnip1', 'Ccne2', 'Pola1', 'Gins
    3', 'Pola2', 'Orc6', 'Prim2', 'Ticrr', 'Orc4', 'Prim1', 'Topbp1', 'Cdk2', 'Ciz1'))
ll <- gmt.obj #unique(gmt.obj$term)
```

do not run again

```
#for (L in names(LL)) {
# group <- toupper(LL[[L]]) #levels(gmt.obj$term)[L]
# nn <- gsub('\\)', '\\(', '\\_', L))
```

```

# gmt_features <- List(group)
# full_obj <- AddModuleScore(
#   object = full_obj,
#   features = gmt_features, ##
#   name = nn,
# )}

namex <- list("DNA_replication_initiation" = "DNA replication initiation",
             "Response_to_DNA_damage_replication_stress" = "Response to DNA
damage replication stress")

p1 <- list()
p <- list()
melt_005df <- list()
melt_005pp <- list()
meltp <- list()
pp <- list()
legend = ''

Idents(full_obj) <- 'Celltype'

#subset <- subset(full_obj, idents='HSC_MPP')
out.dir <- 'Full'
#saveRDS(subset, paste(out.dir, 'HSC_MPP_cluster.rds', sep = '/'))

subset <- readRDS(paste(out.dir, 'HSC_MPP_cluster.rds', sep = '/'))

check_fun <- function(x,df){
  df$comparison <- 'comp'
  for (v in 1:dim(df)[[1]]) {
    d1 <- as.character(x[x$group == as.character(df$Var1[v]), 'treatment']) #
RNA_pop == group
    d2 <- as.character(x[x$group == as.character(df$Var2[v]), 'treatment'])#
RNA_pop == group
    if (d1 == d2) {
      ## keep value in the matrix
      df$comparison <- replace(df$comparison,v,d1)
    }
  }
  return(df)
}

for (i in names(namex)) {
  n <- paste0(i, '1')

```

```

antibio <- data.frame(subset@meta.data[[n]], subset@meta.data$RNA_pop)
antibio$celltype <- 'HSC_MPP'

i <- paste0(i, '_', 'HSC_MPP')
colnames(antibio) <- c('value', 'RNA_pop', 'celltype')
antibio$treatment <- ifelse(antibio$RNA_pop == 'UT_DMSO_pop', 'Not edited',
                           ifelse(antibio$RNA_pop == 'UT_p38i_pop', 'Not
edited',
                                   ifelse(antibio$RNA_pop == 'HS_DMSO_pop', 'HS Cas9',
                                           ifelse(antibio$RNA_pop == 'HS_p38i_pop', 'HS
Cas9',
                                                 ifelse(antibio$RNA_pop == 'GE_DMSO_pop_GFP-',
                                                       'GFP-',
                                                     ifelse(antibio$RNA_pop == 'GE_p38i_pop_GFP-',
                                                           'GFP-',
                                                         ifelse(antibio$RNA_pop == 'GE_DMSO_pop_GFP+',
                                                               'GFP+',
                                                             ifelse(antibio$RNA_pop == 'GE_p38i_pop_GFP+',
                                                                   'GFP+',
                                                                 'ND' )))))))
antibio$treatment <- factor(antibio$treatment, levels=c(
  "Not edited", "HS Cas9", "GFP-", "GFP+"
))

antibio$Sample_name <- ifelse(antibio$RNA_pop == 'UT_DMSO_pop', 'DMSO',
                              ifelse(antibio$RNA_pop == 'UT_p38i_pop', 'p38i',
                                      ifelse(antibio$RNA_pop == 'HS_DMSO_pop', 'DMSO',
                                              ifelse(antibio$RNA_pop == 'HS_p38i_pop', 'p38i',
                                                    ifelse(antibio$RNA_pop == 'GE_DMSO_pop_GFP-',
                                                          'DMSO',
                                                        ifelse(antibio$RNA_pop == 'GE_p38i_pop_GFP-',
                                                            'p38i',
                                                          ifelse(antibio$RNA_pop == 'GE_DMSO_pop_GFP+',
                                                                'DMSO',
                                                            ifelse(antibio$RNA_pop == 'GE_p38i_pop_GFP+',
                                                                'p38i',
                                                              'ND' )))))))

dim(antibio)
alpha_genes <- 0.05
rownames(antibio) <- rownames(subset@meta.data)
colnames(antibio) <- c('value', 'group', 'celltype', 'treatment', 'Sample_name')
## module score and RNA_pop
antibio$group <- factor(antibio$group,
  levels=c("UT_DMSO_pop", "UT_p38i_pop", "HS_DMSO_pop",
           "HS_p38i_pop", "GE_DMSO_pop_GFP+", "GE_p38i_pop_GFP+", "GE_DMSO_pop_GFP-",
           "GE_p38i_pop_GFP-"))

pp[[i]] <- list(pairwise.wilcox.test(antibio$value, antibio$group, p.adjust.method
= "BH", paired = FALSE))
meltpp[[i]] <- data.frame(melt(pp[[i]][[1]]$p.value))

```

```

meltpp[[i]][is.na(meltpp[[i]]$value),3] <- 100 # substitute na third column
if (min(meltpp[[i]]$value) <= alpha_genes){
  melt_005pp[[i]] <- filter(meltpp[[i]] , value <= alpha_genes)

  melt_005df[[i]] <- melt_005pp[[i]][complete.cases(melt_005pp[[i]]),]
  y_position <- list()
  for (y in 1:dim(melt_005df[[i]])[1]) {
    x <- as.numeric(paste(c('0.',y), collapse = ""))
    y_position[y] <- as.numeric(max(antibio$value)+0.1)
  }
  melt_005df[[i]]$y.position <- y_position
  melt_005df[[i]]$p.signif <- symnum(melt_005df[[i]]$value, cutpoints = c(0,
0.0001, 0.001, alpha_genes, 1),
                                     symbols = c( "****", "***", "**", "ns"))

  df <- melt_005df[[i]]
  annotation <- unique(antibio[,c('treatment', 'group')])
  df_x <- check_fun(annotation,df)
  melt_005df[[i]] <- filter(df_x , comparison != "comp")
  if( nrow(melt_005df[[i]]) > 0) {
    ## substitute name

    melt_005df[[i]]$Var1_mod <- ifelse(melt_005df[[i]]$Var1 ==
'UT_DMSO_pop', 'DMSO',
                                     ifelse(melt_005df[[i]]$Var1 ==
'UT_p38i_pop', 'p38i',
                                     ifelse(melt_005df[[i]]$Var1 ==
'HS_DMSO_pop', 'DMSO',
                                     ifelse(melt_005df[[i]]$Var1 == 'HS_p38i_pop',
'p38i',
                                     ifelse(melt_005df[[i]]$Var1 == 'GE_DMSO_pop_GFP-
', 'DMSO',
                                     ifelse(melt_005df[[i]]$Var1 == 'GE_p38i_pop_GFP-
', 'p38i',
                                     ifelse(melt_005df[[i]]$Var1 ==
'GE_DMSO_pop_GFP+', 'DMSO',
                                     ifelse(melt_005df[[i]]$Var1 ==
'GE_p38i_pop_GFP+', 'p38i',
                                     'ND' )))))))

    melt_005df[[i]]$Var2_mod <- ifelse(melt_005df[[i]]$Var2 ==
'UT_DMSO_pop', 'DMSO',
                                     ifelse(melt_005df[[i]]$Var2 ==
'UT_p38i_pop', 'p38i',
                                     ifelse(melt_005df[[i]]$Var2 ==
'HS_DMSO_pop', 'DMSO',
                                     ifelse(melt_005df[[i]]$Var2 == 'HS_p38i_pop',
'p38i',
                                     ifelse(melt_005df[[i]]$Var2 == 'GE_DMSO_pop_GFP-
', 'DMSO',
                                     ifelse(melt_005df[[i]]$Var2 == 'GE_p38i_pop_GFP-

```

```

', 'p38i',
                                     ifelse(melt_005df[[i]]$Var2 ==
'GE_DMSO_pop_GFP+', 'DMSO',
                                     ifelse(melt_005df[[i]]$Var2 ==
'GE_p38i_pop_GFP+', 'p38i',
                                     'ND' )))))))

## comparison df, create table
comparisons <- data.frame(
  .y. = c(rep('len',dim(melt_005df[[i]])[1])),
  group1 = melt_005df[[i]]$Var1_mod, #_mod,
  group2 = melt_005df[[i]]$Var2_mod, #_mod,
  p = melt_005df[[i]]$value,
  p.adj = round(melt_005df[[i]]$value,3),
  p.format = melt_005df[[i]]$value,
  p.signif = melt_005df[[i]]$p.signif,
  method = 'Wc',
  y.position = unlist(melt_005df[[i]]$y.position),
  group = unlist(melt_005df[[i]]$comparison),
  g1 = melt_005df[[i]]$Var1,
  g2 =melt_005df[[i]]$Var2
) %>%
  as.tbl()
#write.table(out_dir, 'DGE_analysis', 'Plot_Lopes', paste0(i, '.txt'),
sep='/')

antibio$str <- antibio$group
p1[[i]] <- ggplot(antibio, aes(x = treatment, y = value, fill=group))+
  geom_split_violin(alpha = .6, trim = FALSE, aes(fill=group)) +
  geom_boxplot(aes(fill=group), outlier.size=0,outlier.alpha =0,
               width = .2, alpha = .7, fatten = NULL, show.legend = FALSE) +
  geom_signif(
    data=comparisons,
    aes(xmin=group, xmax=group, annotations=p.signif, y_position=y.position),
    manual=TRUE, tip_length = 0.,step_increase=0,textsize=4
  )+
  stat_summary(fun.data = "mean_se", geom = "pointrange", show.legend = F,
              position = position_dodge(.175)) +
  theme_minimal() +
  theme(
    legend.position = 'none',
    plot.title = element_text(hjust = 0.5),
    text = element_text(size = 12),
    panel.grid.major = element_blank(),
    panel.grid.minor = element_blank(),
    axis.title.x = element_blank(),
    plot.margin = margin(t = 20, r = 10, b = 10, l = 10, unit = 'pt'),
    axis.line = element_line(colour = "black",
                             size = 0.3, linetype = "solid"),
    axis.text.x = element_text(size=12, color='black'),
    axis.text.y = element_text(size=12, color='black'),

```



```

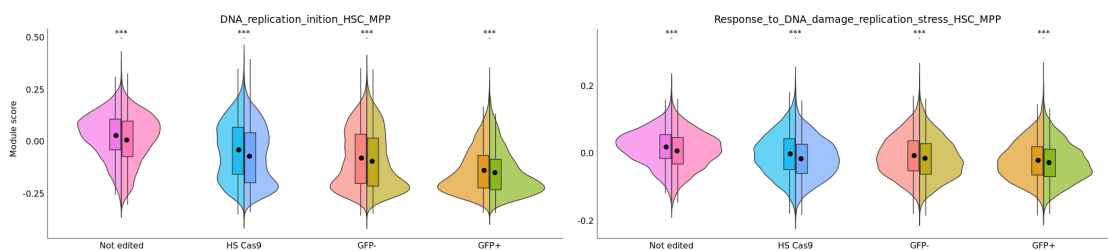
axis.title.y = element_blank(),
strip.placement = "outside") + ggtitle(i)
}
}
}

#Lg_Legend<-function(a.gplot){
# tmp <- ggplot_gtable(ggplot_build(a.gplot))
# leg <- which(sapply(tmp$grobs, function(x) x$name) == "guide-box")
# Legend <- tmp$grobs[[leg]]
# return(Legend)}

#Leg <- get_Legend(p1[[1]])

d <- grid.arrange(grobs = list(p1[[1]],p1[[2]]), left = textGrob("Module score",
rot = 90, vjust = 1),ncol=2)

```



```

#svg(paste(out_dir,'signature_dnarep.svg', sep='/'), width =12, height = 15,
points=12)
#plot(d)
#dev.off()

```

Supplementary Figures

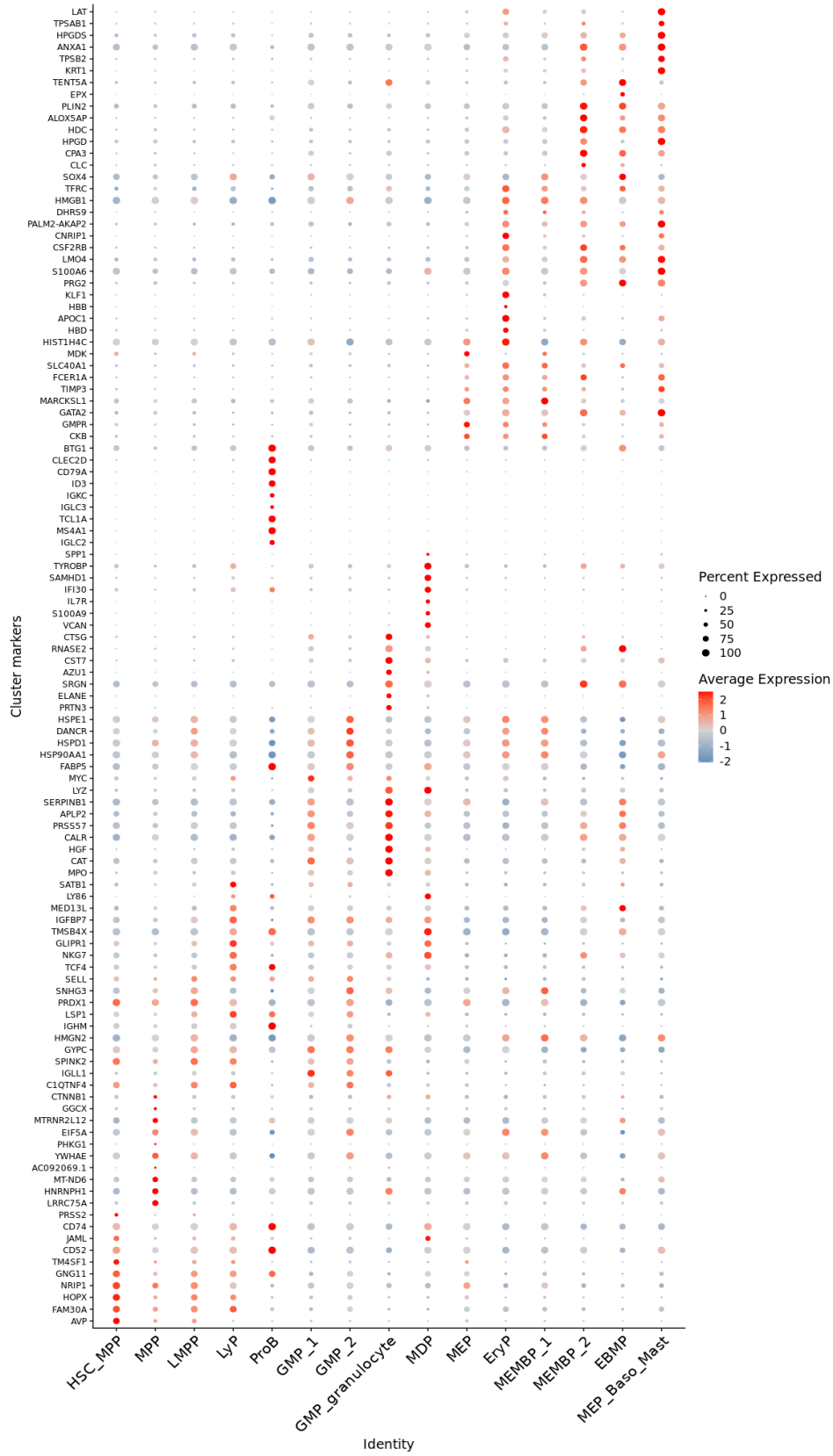
Supplementary Figure 4 a - cluster composition new table and figures

```

Idents(full_obj) <- 'Celltype'
top10 <- full_obj@misc$markers_h$annot_cluster$RNA_snn_h.orig.ident_res.2_refined
%>% group_by(cluster) %>% top_n(n = 10, wt = avg_log2FC)
g <- unique(top10$gene)
d <- DotPlot(object = full_obj, features = g,
             cols = c("white","red"),dot.scale = 3)+ #dot.min
RotatedAxis()+coord_flip()+xlab('Cluster markers')+
theme(axis.text.x=element_text(size=16),axis.text.y=element_text(size=9))+
scale_colour_gradient2(low="steelblue", mid="lightgrey", high="red")+
theme(
  axis.line = element_line(color = "black"),
  axis.ticks = element_line(color = "black"),
  axis.text = element_text(color = "black")
)

```

```
)  
  
#svg(paste(out_dir, "Paper_plots", "Cluster_lineage_markers.svg", sep = "/"), width  
=10, height = 15, pointsize=12)  
#d  
#dev.off()  
d
```



Supplementary figures 4 b - lineages score

```
## Mende et al, Laurenti Lineages
gmt.obj<- read.gmt(paste('reference/GSEA/lineage_hsc_score.gmt',sep='/'))

ll <- unique(gmt.obj$term)

## Calculate module score
# for (L in 1:length(LL)) {
#   group <- LL[L] #Levels(gmt.obj$term)[L]
#   nn <- gsub('\\\',' ',gsub('\(',' ',gsub(' ','_ ',group)))
#   nn <- gsub('-','_ ',nn)
#   glist <- gmt.obj[gmt.obj$term == group,'gene']
#   gmt_features <- list(glist)
#   #LL <- length(x = which(x = gmt_features %in% rownames(x =
full_obj@assays$RNA)))
#   #if (LL != 0){
#     full_obj <- AddModuleScore(
#       object = full_obj,
#       features = gmt_features, ##
#       nbin = 24,
#       ctrl = length(glist), # 100, only for the metabolic pathways
#       seed = 1,
#       name = nn,
#       search = TRUE
#     )
#   }

#saveRDS(full_obj, paste(out_dir, 'Full_final_v7.rds', sep = '/'))

## GMP score
Idents(full_obj) <- 'Celltype'
l <- unique(gmt.obj$term)
p <- list()
for (t in l) {
  n <- gsub('[.]', '\n',t)
  p[[t]] <- VlnPlot(object = full_obj, features = paste0(t, "1"), #group.by =
c("CSC_clusters"),
  pt.size = 0,sort = F,col = mycols)+
  geom_boxplot(width=0.2,outlier.shape = NA)+ ylab(n)+
  theme(legend.position="none",title = element_blank(),
  axis.title.x = element_blank(),
  axis.text=element_text(size=16),
  axis.line = element_line(colour = "black", size=0.5),
  axis.title.y = element_text(size=16),
  axis.text.x = element_text(hjust=1,size=16),
  axis.text.y = element_text(size = 16),
  panel.grid = element_blank(),
  panel.border = element_blank(),
  text = element_text( size= 16))+ggtitle('')
```

```

}
## get Legend just from one
lg_legend<-function(a.gplot){
  tmp <- ggplot_gtable(ggplot_build(a.gplot))
  leg <- which(sapply(tmp$grobs, function(x) x$name) == "guide-box")
  legend <- tmp$grobs[[leg]]
  return(legend)}

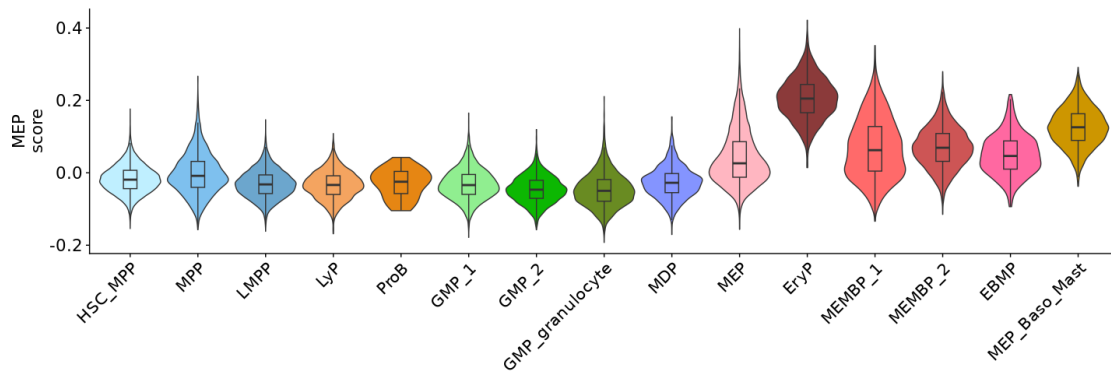
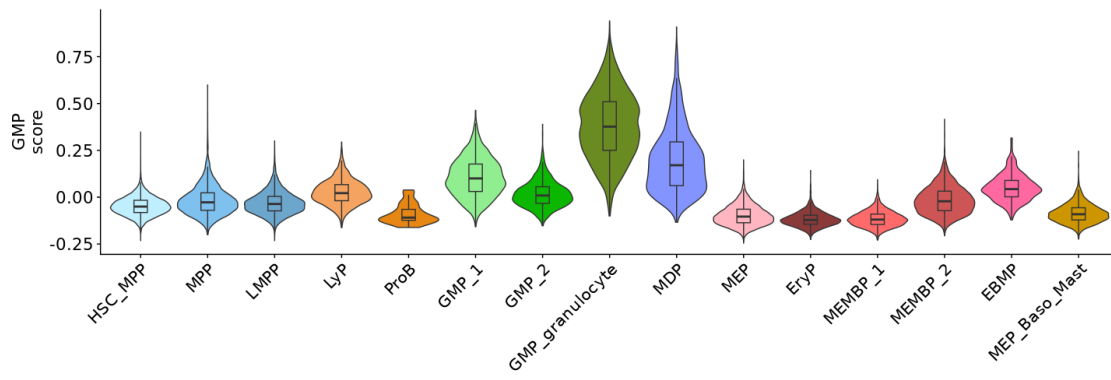
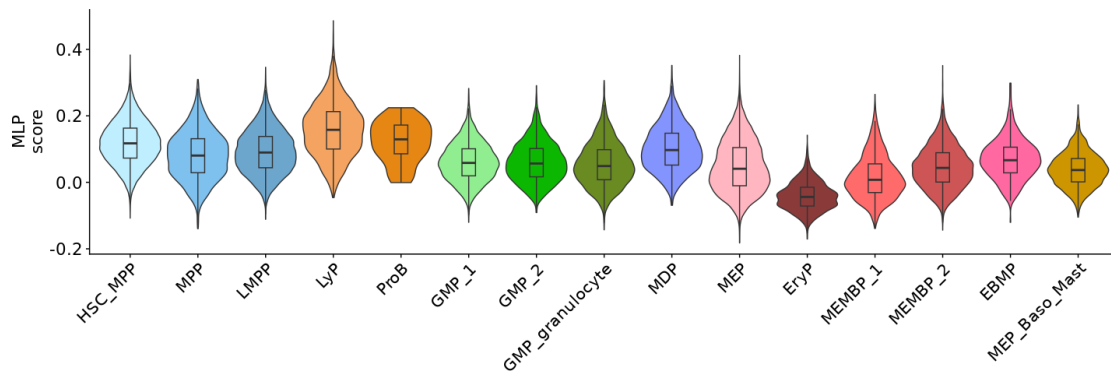
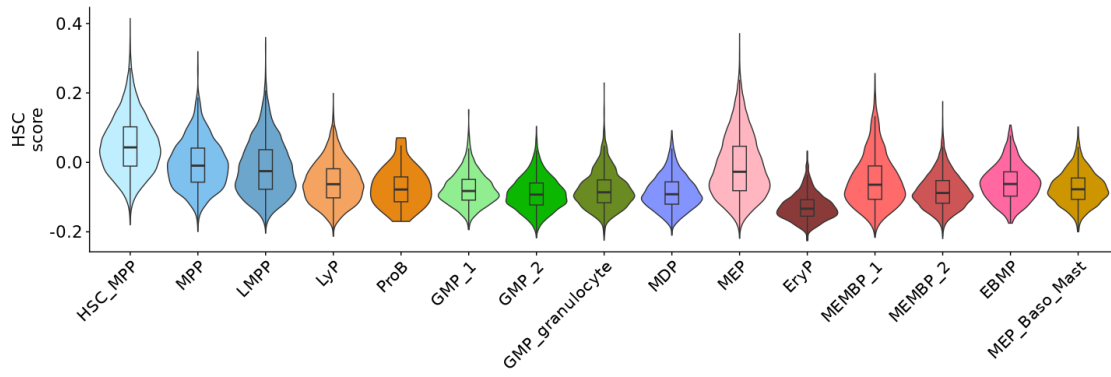
legend <- VlnPlot(object = full_obj, features = paste0(t, "1"), #group.by =
c("CSC_clusters"),
                 pt.size = 0,sort = TRUE, col = mycols)+
  geom_boxplot(width=0.2,outlier.shape = NA)

mylegend<- lg_legend(legend)

#svg(paste(out_dir,"Paper_plots", "Lineage_scores_annotated_nonsorted.svg",
sep='/'), width =8, height = 15, pointsize=12)
#grid.arrange(p$HSC.score, p$MLP.score, p$GMP.score,p$MEP.score, #myLegend,
#            nrow = 4,top = textGrob("",gp=gpar(fontsize=20,font=3)))
#dev.off()

grid.arrange(p$HSC.score, p$MLP.score, p$GMP.score,p$MEP.score, #myLegend,
            nrow = 4,top = textGrob("",gp=gpar(fontsize=20,font=3)))

```



Supplementary figures 4 c - gsea plot

```
hallmark.subset <- NULL
hallmark.subset <-
read.table('Full/Paper_plots/tables/GSEA_results_p38_sig_sel.txt')

min0 <- min(hallmark.subset[!is.na(hallmark.subset$NES), 'NES'])
max0 <- max(hallmark.subset[!is.na(hallmark.subset$NES), 'NES'])

hallmark.subset$stars=stars.pval(hallmark.subset$pvalue)

hallmark.subset$ID <- str_wrap(hallmark.subset$ID ,width = 45)
hallmark.subset$Name <- as.character(hallmark.subset$ID)
hallmark.subset$celltype <- gsub('_', ' ', hallmark.subset$celltype)
hallmark.subset$celltype <- ifelse(hallmark.subset$celltype == 'MEP Baso Mast',
'MEP Baso-Mast', hallmark.subset$celltype)

hallmark.subset$celltype <- factor(hallmark.subset$celltype, levels =
c("HSC MPP", "MPP", "LMPP", "GMP 1",
"GMP 2", "GMP granulocyte",
"MDP", "MEP", "EryP", "MEMBP 1",
"MEMBP 2", "MEP Baso-
Mast", "LyP", "ProB", "EBMP" ))

hallmark.order <- hallmark.subset%>% group_by(ID) %>% summarise(Pos = sum(NES))
hallmark.order.terms <- hallmark.order[order(hallmark.order$Pos, decreasing =
TRUE), "ID", drop = FALSE]
hallmark.subset$ID <- factor(hallmark.subset$ID, levels = hallmark.order.terms$ID)

hallmark.subset$Dataset <- factor(hallmark.subset$Dataset,
levels=c("UT_p38i_pop_vs_UT_DMSO_pop",
"HS_p38i_pop_vs_HS_DMSO_pop",
"GE_p38i_pop_GFPneg_vs_GE_DMSO_pop_GFPneg",
"GE_p38i_pop_GFPpos_vs_GE_DMSO_pop_GFPpos" ) )

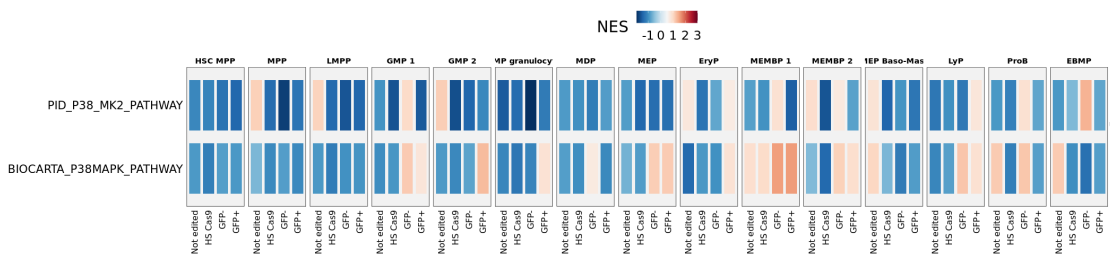
hallmark.subset$Dataset <- ifelse(hallmark.subset$Dataset ==
"UT_p38i_pop_vs_UT_DMSO_pop", 'Not edited',
ifelse(hallmark.subset$Dataset ==
"HS_p38i_pop_vs_HS_DMSO_pop", 'HS Cas9',
ifelse(hallmark.subset$Dataset ==
"GE_p38i_pop_GFPneg_vs_GE_DMSO_pop_GFPneg", "GFP-",
ifelse(hallmark.subset$Dataset ==
"GE_p38i_pop_GFPpos_vs_GE_DMSO_pop_GFPpos", "GFP+", 'none' ))))

hallmark.subset$Dataset <- factor(hallmark.subset$Dataset, levels=c('Not edited',
```

```
'HS Cas9',
"GFP-",
"GFP+")
```

```
p_p38 <- ggplot(data = hallmark.subset, mapping = aes(x = Dataset, y =
reorder(Name, -NES))) +
  geom_tile(aes(fill=NES, group = celltype), width=0.80, height=0.80)+
  facet_grid(Category ~ celltype , scales = "free_y", space = "free", )+
  theme(text= element_text(size=22,color='black'),
        axis.text.x = element_text(angle = 90, vjust = 0.5, hjust=1,
color='black', size =12),
        axis.text.y = element_text(color='black', size =16),
        plot.title = element_text(hjust = 0), plot.subtitle = element_text(hjust =
0),
        axis.ticks = element_blank(), axis.text = element_text(size = 8),
axis.title = element_blank(),
        panel.grid.major = element_blank(), panel.grid.minor = element_blank(),
        panel.background = element_rect(fill = "gray95", color='black'),
        strip.background = element_rect(colour="white",
fill="white",size=0.5),
        strip.text = element_text(color='black',face = 'bold', size = 11),
        legend.position = 'top', legend.direction = 'horizontal') +
  scale_fill_gradientn(colours =
colorRampPalette(rev(brewer.pal(11,"RdBu")))(100),
        limits = c(min0,3),
        na.value = "grey")

#wdir='Full/DGE_analysis'
#svg(paste(wdir,"HM_categories_v3_P38_pid.svg",sep = '/'),width=25,
# height=3, pointsize=12)
#p
#dev.off()
p_p38
```



Supplementary figures 4 e - Lopes et al genes

```
##### plot gene for each cluster and for each condition
c <- list("UT_p38i_pop_vs_UT_DMSO_pop" = c("UT_p38i_pop", "UT_DMSO_pop"),
```



```

      "HS_p38i_pop_vs_HS_DMSO_pop" = c( "HS_p38i_pop", "HS_DMSO_pop"),
      "GE_DMSO_pop_GFP+_vs_GE_DMSO_pop_GFP-" =
c("GE_DMSO_pop_GFP+", "GE_DMSO_pop_GFP-"), ## h3 vs h3
      "GE_p38i_pop_GFP+_vs_GE_DMSO_pop_GFP+" =
c("GE_p38i_pop_GFP+", "GE_DMSO_pop_GFP+"), ## h4 vs h3
      "GE_p38i_pop_GFP-_vs_GE_DMSO_pop_GFP-" = c("GE_p38i_pop_GFP-
", "GE_DMSO_pop_GFP-"),
      "GE_DMSO_pop_GFP+_vs_UT_DMSO_pop" = c("GE_DMSO_pop_GFP+", "UT_DMSO_pop"),
## h3 vs h3
      "GE_DMSO_pop_GFP-_vs_UT_DMSO_pop" = c("GE_DMSO_pop_GFP-", "UT_DMSO_pop"),
      "GE_p38i_pop_GFP+_vs_UT_DMSO_pop" = c("GE_p38i_pop_GFP+", "UT_DMSO_pop"),
      "GE_p38i_pop_GFP-_vs_UT_DMSO_pop" = c("GE_p38i_pop_GFP-", "UT_DMSO_pop")
## h3 vs h3

## dge analysis tables
marker_dir <- 'Full/DGE_analysis/DGE_alivecd90_together/tables'

Idents(full_obj) <- full_obj$Celltype

x <- unique(full_obj$Celltype)
x <- c('MEMBP_2', 'LMPP', 'LyP', 'HSC_MPP', 'EryP',
      'MEP', 'GMP_2', 'MEMBP_1', 'MPP', 'GMP_granulocyte', 'MEP_Baso_Mast',
      'GMP_1', 'MDP', 'EBMP', 'ProB' )
xnew <- paste0(as.character(x), '_')

cnew <- paste0('_', as.character(names(c)))
cnew <- gsub('\\+', 'pos', cnew)
cnew <- gsub('\\-', 'neg', cnew)

comparisons <- expand.grid(x, names(c))
comp <- paste0(comparisons$Var1, '_', comparisons$Var2)
markers_all <- data.frame()
for (comp in comp) { #for each comparisons within a cluster
  #cat(comp, '\n')
  ### get name comparison and celltype separated
  compx_original <- compx
  compx <- gsub('\\+', 'pos', compx)
  compx <- gsub('\\-', 'neg', compx)
  celltype <- unique(str_replace(compx, cnew, ''))

  cp <- celltype[order(nchar(celltype), celltype)][1] ## cell type

  nx <- unique(str_replace_all(compx, as.character(xnew), ''))
  n <- nx[order(nchar(nx), nx)][1] ## comparisons
  f <- paste(marker_dir, paste0(compx_original, '.txt'), sep = '/')

  if (file.exists(f)) {
    markers <- read.table(f)
  }
}

```

```

markers$Celltype <- cp
markers$Contrast <- n
markers$Comparisons <- compx
markers$gene <- rownames(markers)
rownames(markers) <- NULL
}
if (nrow(markers_all) == 0) {
  markers_all <- markers
} else {
  markers_all <- rbind(markers_all, markers)
}
}
rownames(markers_all) <- NULL

markers_all <- markers_all[order(markers_all$avg_log2FC),]

genes <- list('FORK_REVERSAL'=c('FBH1', 'SMARCAL1', 'ZRANB3', 'HLTF'),
             'FORK_RESTART' = c('RECQL', 'WRN', 'RNF8', 'DNA2'),
             'REPRIMING' = c('BLM', 'RMI1', 'TOP3A', 'RMI2', 'PRIMPOL'))

markers_all_genes <- markers_all
hsc_genes <- markers_all_genes[markers_all_genes$gene %in% unname(unlist(genes)) &
markers_all_genes$Celltype == 'HSC_MPP',]

#print(hsc_genes[order(hsc_genes$avg_log2FC, decreasing = FALSE), ] )

markers_all_subset <- hsc_genes

markers_all_subset$pvalue=stars.pval(markers_all_subset$p_val_adj)

markers_all_subset$Category <- ifelse(markers_all_subset$gene %in%
genes$FORK_REVERSAL, 'FORK_REVERSAL',
                                     ifelse(markers_all_subset$gene %in%
genes$FORK_RESTART, 'FORK_RESTART',
                                             'REPRIMING'))

min0 <- min(markers_all_subset$avg_log2FC)
max0 <- max(markers_all_subset$avg_log2FC)
markers_all_subset <- markers_all_subset[markers_all_subset$Contrast %in%
c("UT_p38i_pop_vs_UT_DMSO_pop", "GE_p38i_pop_GFPpos_vs_GE_DMSO_pop_GFPpos",
"GE_p38i_pop_GFPneg_vs_GE_DMSO_pop_GFPneg", "HS_p38i_pop_vs_HS_DMSO_pop" ),]

markers_all_subset$treatment <- ifelse(markers_all_subset$Contrast ==
"UT_p38i_pop_vs_UT_DMSO_pop", 'Not edited',
                                       ifelse(markers_all_subset$Contrast ==
"HS_p38i_pop_vs_HS_DMSO_pop", 'HS Cas9',

```

```

        ifelse(markers_all_subset$Contrast ==
"GE_p38i_pop_GFPneg_vs_GE_DMSO_pop_GFPneg", 'GFP-',
        ifelse(markers_all_subset$Contrast ==
"GE_p38i_pop_GFPpos_vs_GE_DMSO_pop_GFPpos", 'GFP+', 'nd'))))

markers_all_subset$treatment <- factor(markers_all_subset$treatment, levels=c('Not
edited', 'HS Cas9', "GFP-", "GFP+"))
markers_all_subset$Celltype <- ifelse(markers_all_subset$Celltype == 'CMP',
'MPP', markers_all_subset$Celltype)
markers_all_subset$Celltype <- factor( markers_all_subset$Celltype, levels =
levels(full_obj$Celltype))

hsc_plot <- ggplot(markers_all_subset, aes(y = reorder(gene, -avg_log2FC), x
=treatment, fill=avg_log2FC)) +
  geom_tile(aes(fill=avg_log2FC, group = Celltype), width=0.80, height=0.80)+
  facet_grid(Category ~ Celltype , scales = "free_y", space = "free", )+
  theme(text= element_text(size=22,color='black'),
        axis.text.x = element_text(angle = 90, vjust = 1, hjust =1, color='black',
size =12),
        axis.text.y = element_text(color='black', size =10),
        plot.title = element_text(hjust = 0), plot.subtitle = element_text(hjust =
0),
        axis.ticks = element_blank(), axis.text = element_text(size = 8),
axis.title = element_blank(),
        panel.grid.major = element_blank(), panel.grid.minor = element_blank(),
        panel.background = element_rect(fill = "gray95", color='black'),
        strip.background = element_rect(colour="white",
        fill="white",size=0.5),
        strip.text = element_text(color='black',face = 'bold', size = 11)) +
  geom_text(aes(label = pvalue),size =5) +
  scale_fill_gradientn(colours = c('blue', 'white', 'red'),
        na.value = "grey",
        rescaler = ~ scales::rescale_mid(.x, mid = 0))+
  ggtitle('p38i vs DMSO')

#svg(paste('Full/DGE_analysis', "DGE_Lopes_genes.svg", sep = '/'),width=6,
# height=12, pointsize=12)
#hsc_plot
#dev.off()
hsc_plot

```

p38i vs DMSO

